METHODS OF TREATMENT OF INFLAMMATORY DISEASES

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A01N 37/36

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Field of Classification Search ........ 514/159–166; 514/474

See application file for complete search history.

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(Continued)

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ABSTRACT

Inflammatory bowel diseases are represented by two idiopathic disorders, which include ulcerative colitis and Crohn’s disease. Ulcerative colitis is restricted to the colon and involves uncertain and inflammation of the lining (mucosa) of the large intestine. Crohn’s disease, on the other hand, can involve the mucosa of the small and/or large intestine and may involve deeper layers of the bowel wall. The present invention contains N-acetylcysteine disposed in a pharmaceutically acceptable carrier for oral or rectal delivery for treating such inflammatory bowel diseases by topical application within the colon.

13 Claims, 6 Drawing Sheets
OTHER PUBLICATIONS


* cited by examiner
Figure 4

Aggregate Microscopic Injury

Injury Score

p < 0.05

ASA/NAC

NAC

5-ASA

TNBS
METHODS OF TREATMENT OF INFLAMMATORY DISEASES

This application is a continuation of U.S. Ser. No. 11/023, 812, now U.S. Pat. No. 7,417,037, filed Dec. 28, 2004 and issued Aug. 26, 2008, and also claims the benefit under 35 U.S.C. 119(e) of U.S. Provisional Application Ser. No. 60/537,766, filed Jan. 20, 2004, the entire disclosure of both of which is hereby expressly incorporated by reference herein.

BACKGROUND

Inflammatory bowel diseases (IBDs) including ulcerative colitis and Crohn’s disease, are complex diseases that are thought to result from a combination of genetic and environmental factors (10). The discovery of the adaptive immune response in the gut mucosa has led to a better understanding of the pathogenesis of IBD (11). The role of the gut microbiome in the pathogenesis of IBD is also gaining increasing attention (12).

Antimicrobial agents (AMAs) have been used to treat IBD, but their effectiveness is limited. The use of 5-aminosalicylic acid (5-ASA) has been shown to be effective in the treatment of IBD (13, 14). However, the use of 5-ASA is limited by its side effects, such as diarrhea and abdominal cramping (15).

5-ASA agents are presently available for the treatment of mild to moderate inflammatory bowel disease. Recent investigations into the etiologic triad of genetic: environmental: immune factors have expanded our knowledge of the role of these individual components and their potential interactions. Pathologic studies of IBD envision initiating events, possible interactions, arising from the gut lumen that converts immune tolerance to a sustained hyperactive state with elaboration and amplification of cellular and humoral mediators. Immune response derived injurious and proinflammatory substances cause tissue injury and destruction. These substances include prostaglandins, reactive oxygen metabolites, nitric oxide, leukotrienes, proteases and matrix metalloproteinases (3). The role of reactive oxygen species (ROS) and nitric oxide (NO) have been examined in experimental models of IBD (4, 5). Pharmacological and genetic manipulation of oxygen free radical and NO generation have been shown to accelerate experimental colitis induced by luminal administered trinitrotoluene sulfoonic acid (TNBS) and dextran sulfate sodium (DSS) (5, 10).

Although several experimental strategies have been employed that suggest the importance of enhanced production of superoxide and nitric oxide in the pathogenesis of IBD, inconsistent results have the issue unresolved. For example, the beneficial effect of superoxide dismutase (SOD) treatment in experimental models of colitis has been reported while SOD treatment in humans with IBD has shown limited benefit (40). Similarly, the inhibitors of inducible nitric oxide synthase (iNOS) have yielded mixed results in various experimental models of IBD (6-8, 27). The antioxidants N-acetylcysteine (NAC) and phenyl N-tert-butylnitrone (PBN) when used alone have shown to be effective in protection against TNBS-induced colitis in rat (9) and DSS-induced colitis in mice, (10) respectively. Furthermore, recent studies suggest a dominant role of iNOS-derived NO in a murine model of colitis (5). Antioxidant therapy has also been shown to suppress colonic iNOS activity and to decrease colonic NF-kB DNA-binding activity in experimental animals (10). Nuclear factor-kB, NF-kB is a family of transcription factors known to regulate a variety of genes controlling the inflammatory process and regulating programmed cell death (41).

Thus, there exists extensive experimental support for the notion that reactive oxygen molecules and nitric oxide may contribute to the pathogenesis of mucosal injury in inflammatory bowel disease. Furthermore, experimental evidence also provides support for the concept that inhibition of nitric oxide species and NO generation exert favorable effects on mucosal healing and the inflammatory process in several well-defined models of chemically induced colitis. However, there continues to be a need in the field for a more effective treatment of inflammatory bowel diseases and other conditions related to inflammation. It is to this need that the present invention is directed.

SUMMARY OF THE INVENTION

Disclosed herein are methods of treating an inflammatory bowel disease comprising administering a composition comprising N-acetylcysteine or a combination of 5-aminosalicylic acid and N-acetylcysteine to a mammal or contacting a colonic cell of the mammal with the said composition. Also disclosed are methods of reducing cytokine gene expression or reducing myeloperoxidase activity in colonic tissue of a mammal, the method comprising administering a composition comprising 5-aminosalicylic acid and N-acetylcysteine to a mammal or contacting a colonic cell of the mammal with the said composition.
Both 5-ASA and antioxidants such as NAC and phenyl N-tert-butylnitroene have relatively low profiles for toxicity. 5-ASA may be associated with allergic reactions to the medications and should be avoided in patients with aspirin sensitivity. As with any nonsteroidal agent there exists potential for hepatic and renal toxicity. N-acetylcysteine or Mucomyst™ has had a wide experience in man for the treatment of acetylaminophen hepatotoxicity. This agent has proven safe. Other antioxidants contemplated for use herein in conjunction with 5-ASA include other aminosuglycates including 4-aminosalicylic acid (4-ASA), and N-acetyl-5-aminosalicylic acid; other nonsteroidal anti-inflammatory drugs (NSALDs) including those that inhibit cyclooxgenase I and/or II, such as sulindac, celecoxib and refucoxib; ascorbate; vitamin C; vitamin A; Vitamin E; beta-carotene; herbal agents such as milk thistle; selenium; iron in various ferric and ferrous formulations; phospholipase A2 inhibitors, e.g., carboxymethylcellulose-linked phosphatidylethanolamine; superoxide dismutase mimetics, such as Mn/III tetrakis (1-methyl-4-piperidyl) of MnTMPyP; melatonin; zolmitriptan; rebamipide; and phenyl N-tert-butylnitroene (PBN); and combinations of any of the above.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1: Experimental Design: Colitis was induced at day 0 by rectal administration of TNBS. Three days after TNBS dosing rats received daily intraluminal therapy per rectum for either 5 or 8 days. Four treatment protocols were employed and included: (A) 5-ASA (100 mg/kg), (B) NAC (40 mM), (C) a combination of 5-ASA (100 mg/kg) and NAC (40 mM), and (D) a saline control.

FIG. 2: Macroscopic grading of inflammation and injury 8 days after intracolonic administration of TNBS. Macroscopic grading (0-6) 8 days after TNBS revealed moderately severe inflammation and ulceration: score 4.5±0.5. Therapy with the NAC/5-ASA combination for 5 df was the only intervention that significantly reduced macroscopic injury: score 2.6±0.7. These results are representative of 4 different experiments done at different times with 5 animals per group per each experiment. * denotes p<0.05 vs TNBS.

FIG. 3: Macroscopic injury score 11 days after TNBS and 8 days of treatment with 5-ASA alone, NAC alone and the NAC plus 5-ASA combination.

FIG. 4: Microscopic grading of inflammation and injury 8 days after intracolonic administration of TNBS. Microscopically, TNBS affected significantly each parameter of injury and inflammation as reflected by a cumulative microscopic injury score (0-15) of 9.4±1.0 p<0.001. The NAC/5-ASA combination significantly reduced the cumulative microscopic injury: score 5.0±1.2 p<0.001. Single agent therapy with either NAC alone or 5-ASA alone did not significantly decrease microscopic injury: NAC 9.0±1.8, and 5-ASA 9.0±0.9. These results are representative of 4 different experiments done at different times with 5 animals per group per experiment. * denotes p<0.05 vs TNBS.

FIG. 5: Combination therapy with the NAC/5-ASA combination for 8 days reduced significantly the aggregate microscopic injury score by 75%. Therapy was initiated 3 days after induction of colitis by TNBS. Furthermore, single agent therapy with 5-ASA alone and NAC alone also significantly decreased histological measures of injury by at least 46 and 53%, respectively. The degree of histologic healing with the NAC/5-ASA combination was significantly greater than that observed with either 5-ASA or NAC alone.

FIG. 6: Myeloperoxidase activity in colon tissue 11 days after TNBS and 8 days of treatment with 5-ASA alone, NAC alone and the NAC plus 5-ASA combination. Results represent the means±SEM of 4-6 rats per condition.

DESCRIPTION OF THE INVENTION

The present invention contemplates use of 5-ASA (mesalamine) plus an antioxidant either together as separate molecular entities or when coupled chemically, to provide an enhanced therapeutic or prophylactic effect against inflammatory bowel diseases in mammals. The combination of mesalamine plus an antioxidant in a mammal model of colitis promotes healing and reduces inflammation to a significantly greater degree than either agent used alone. The present invention provides an improved anti-inflammatory effectiveness of 5-ASA for use in the treatment of inflammatory bowel disease by the addition of an antioxidant. Use of the presently claimed compositions result in a reduction in the need for additional anti-inflammatory agents, such as prednisone and Ibruln in the treatment of inflammatory disease (thereby providing a significant cost benefit and reduction in drug-induced toxicity). Furthermore, the present invention can serve as a model for additional pharmacological approaches to other inflammatory and, possibly, neoplastic conditions within the gastrointestinal tract.

As shown below experiments were performed to examine the ability of an antioxidant (NAC) and 5-ASA, when administered separately or in combination to the distal colon, to affect mucosal healing and repair following TNBS or DSS.

Materials and Methods

Experimental Animals

Male Sprague Dawley rats (200-250 grams) were housed in cages containing contact bedding. Rats were deprived of food for 24 hrs prior to the induction of colitis, but were allowed free access to water throughout the experiment. Institutional approval for experimental protocols was provided by the research and animal care committees of the research services at the Oklahoma City Veterans Administration Medical Center.

Induction of Colitis by TNBS Treatment

Following a 24 hr fast, rats were lightly anesthetized using isoflurane followed by insertion of a polyethylene catheter into the anus. The tip of the catheter was advanced 8 cm proximal from the anus and a single dose of TNBS (Sigma, St. Louis, Mo.), dissolved in 50% ethanol, was injected through the cannula (15 mg dissolved in a 0.6 ml volume of 50% ethanol) to induce colitis. Following the administration of TNBS the animals were maintained in a head-down position for approximately 60 seconds to prevent leakage of the instillate. After 72 hrs, rats dosed initially with TNBS were randomly assigned to one of four treatment groups to receive daily intracolonic therapy in a manner similar to that used for TNBS administration. The groups were: A) 5-aminosalicylic acid (5-ASA) (100 mg/kg); B) N-acetylcysteine (NAC) (40 mM); C) 5-ASA (100 mg/kg) plus NAC (40 mM); or D) saline. An additional group of rats (group E) served as control and received saline per rectum on day 0 and at subsequent intervals that corresponded to treatment protocols. Rats were treated for either 5 or 8 days and then sacrificed by cervical dislocation. FIG. 1 illustrates the design of these experiments.

Induction of Colitis by DSS Treatment

Colitis was induced by oral administration of a 4% solution of DSS (dextran sulfate sodium) in drinking water for 4 days. Experimental groups were randomized to include group 1 control, group 2-5 received DSS in drinking water and groups 3-5 were further randomized to receive daily intracolonic treatment with 5-ASA (100 mg/kg) plus NAC (40 mM),
group 3; 5-ASA alone, group 4; and NAC constituted group 5. Animals received treatment for 4 days. Macroscopic indices of colonic injury were scored and tabulated. Histological features that were examined included epithelial damage and mucosal ulceration. Additional measures included determination of serum C-reactive protein (CRP) levels and cytokine gene expression in colonic tissues performed by ribonuclease protection assay (RPA).

Macroscopic Assessment of Colitis

Upon sacrifice, the distal 8 cm of the colon was removed, opened by a longitudinal incision and rinsed with phosphate buffered saline to remove fecal material. Macroscopic assessment of colitis was carried out by an independent observer who was unaware of the treatment groups. The criterial and scale of grading are listed in Table 1. Inflammation was present if the mucosa was erythematous. Ulceration of the mucosa was defined as a distinct break or interruption of the mucosa. Once macroscopic damage was assessed full thickness colonic tissue samples were taken from the inflamed areas and either processed for histology or snap frozen in liquid nitrogen for subsequent cytokine measurement and determination of myeloperoxidase (MPO) activity.

### TABLE 1

<table>
<thead>
<tr>
<th>Macroscopic Injury</th>
<th>Score</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>0</td>
</tr>
<tr>
<td>One area of inflammation</td>
<td>1</td>
</tr>
<tr>
<td>No ulcer</td>
<td></td>
</tr>
<tr>
<td>No inflammation</td>
<td></td>
</tr>
<tr>
<td>One ulcer</td>
<td>2</td>
</tr>
<tr>
<td>One area of inflammation</td>
<td>3</td>
</tr>
<tr>
<td>One or two ulcers</td>
<td></td>
</tr>
<tr>
<td>One area of inflammation</td>
<td>4</td>
</tr>
<tr>
<td>More than 2 ulcers</td>
<td></td>
</tr>
<tr>
<td>Two areas of inflammation</td>
<td>5</td>
</tr>
<tr>
<td>More than 2 ulcers</td>
<td></td>
</tr>
<tr>
<td>Ulceration &gt;2 cm</td>
<td>6</td>
</tr>
</tbody>
</table>

Histological Assessment of Colitis

Colonic tissue samples taken from the initial segment were immersed in 10% phosphate buffered formalin and subsequently embedded in paraffin. Sections of 3 micron thickness were cut and stained with hematoxylin and eosin. The slides were then evaluated by a pathologist for epithelial damage, architectural changes, mononuclear infiltration, polymorphonuclear leukocyte (PMN) infiltration and ulceration. The individual microscopic features of colitis were graded according to criteria noted in Table 2. In addition to scoring individual features of colitis, an aggregate score of colitis was tabulated by adding together individual scores, thus, providing a global assessment of colitis.

### TABLE 2

<table>
<thead>
<tr>
<th>Histological Scoring of Colitis</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Epithelial Damage</strong></td>
</tr>
<tr>
<td>0 Normal</td>
</tr>
<tr>
<td>1 Focal Mucosal injury</td>
</tr>
<tr>
<td>2 Extensive mucosal injury</td>
</tr>
<tr>
<td><strong>Architectural Damage</strong></td>
</tr>
<tr>
<td>0 Normal</td>
</tr>
<tr>
<td>1 Moderately disturbed</td>
</tr>
<tr>
<td>2 Severely disturbed</td>
</tr>
<tr>
<td><strong>Mononuclear Infiltration</strong></td>
</tr>
<tr>
<td>0 Normal</td>
</tr>
<tr>
<td>1 Moderate increase</td>
</tr>
<tr>
<td>2 Severe increase</td>
</tr>
<tr>
<td><strong>PMN Infiltration</strong></td>
</tr>
<tr>
<td>0 Normal</td>
</tr>
<tr>
<td>1 Present in surface</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Histological Scoring of Colitis</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Ulcerations</strong></td>
</tr>
<tr>
<td>0 None</td>
</tr>
<tr>
<td>1 1-33% Ulcerated</td>
</tr>
<tr>
<td>2 34%-66% Ulcerated</td>
</tr>
<tr>
<td>3 67% Ulcerated</td>
</tr>
</tbody>
</table>

Myeloperoxidase Activity

Myeloperoxidase activity was used as an indirect measure of the severity of colonic inflammation by PMNs. Whole thickness tissues taken following macroscopic assessment were weighed (100 mg) and immediately snap frozen in liquid nitrogen for storage at 80°C. The tissues were then removed from storage to 80°C and allowed to thaw on ice. Once thawed, 1 ml of hexadecyltrimethylammonium bromide (HTAB) (Sigma, St. Louis, Mo.) containing 50 mM KH₂PO₄ (Sigma, St. Louis, Mo.) and 0.1M Na₂HPO₄ (Sigma, St. Louis, Mo.) was added per 100 mg tissue for homogenization. Homogenates then underwent a series of four freeze/thaw cycles before finally being centrifuged at 12,000 x g for 10 minutes at 4°C. The supernatant was collected for measurement of MPO activity. Horseradish peroxidase (Sigma, St. Louis, Mo.) was used as a standard; stock solution of 0.5 mg/ml Tetramethylbenzidine (TMB) (Sigma, St. Louis, Mo.) was used as the substrate for carrying out the reaction. At the time of assay 25 µl of standard and sample were added to appropriately labeled tubes. TMB was added at a volume of 250 µl to initiate the reaction and 0.1 M H₂SO₄ (250 µl) was added after 10 minutes to terminate the reaction. The absorbance changes were read at 450 nm and recorded. Results were expressed as nm/g of tissue.

Ribonuclease Protection Assay (RPA)

Following the manufacturer’s protocol, 1 ml of TRI Reagent (Sigma, St. Louis, Mo.) is used per 100 mg of tissue. The frozen tissue is placed into the TRI Reagent and immediately homogenized using a tissue macerator. The homogenates are transferred to microcentrifuge tubes and 0.2 ml of chloroform is added per mL of TRI Reagent used. The samples are shaken thoroughly and allowed to sit at room temp for 3 min. Samples were then centrifuged at 12,000 x g for 15 min at 4°C. After centrifugation, the supernates were transferred to a new tube and 9.5 ml of 2-propanol is added to precipitate the DNA. After sitting at room temp for 10 min, the samples are centrifuged at 12,000 x g for 15 min. The supernates are discarded and the pellets are washed in 70% ethanol for 5 min. The ethanol is removed and the pellets are dissolved in 50 µl of DEPC treated water per sample. Quantitation is performed by spectrophotometry at 260 nm and 280 nm.

RPA analysis was performed with RiboQuant™ multi-probe RNase protection assay system (BD Biosciences-Pharmingen, San Diego, Calif.). To 20 µg of total RNA in 8 µl of hybridization buffer and 2 µl of 32P-labeled in vitro transcribed RNA probes were added. The probes were transcribed from the multiprobe template set rCK-1 which as a panel of probes against IL-1α, IL-1β, IL-3, IL-5, IL-10, IL-2, TNFα, TNFβ, GADPH genes. Probes were prepared according to the manufacturer's protocol. The RNA samples containing labeled probes were incubated briefly at 90°C, slowly cooled to 50°C and hybridized overnight at that temperature. The samples were then slowly cooled to 30°C. Thereafter, a mixture of RNases A and T1 was added to digest single-
stranded RNA leaving the double-stranded RNA formed by the probes annealing to their cognate mRNAs intact. Following proteinase K digestion and phenol-chloroform extraction, the undigested double-stranded RNA was precipitated with ammonium acetate and ethanol. After centrifugation at 14,000×g for 20 min, the pellets are washed with 90% ethanol and recentrifuged. The ethanol was removed and the pellets were dissolved in 5 μl of formamide loading dye. The double-stranded RNAs were resolved on a 5% polyacryla-

mide/urea gel run in TBE buffer. After electrophoresis, the gel was dried at 80°C under vacuum and then exposed to a PhosphorImager screen overnight. Detection and quantitation of the resolved bands on the gel were performed on a Storm PhosphorImager™ system (Amersham Biosciences, Piscataway, N.J.).

Statistical Significance

All values in the figures and text are expressed as mean±standard error of the mean (SEM). The statistical significance of any difference among groups was analyzed using Student’s two-tailed t test for equal and unequal variance observations. P values of <0.05 were considered to be statistically significant.

Results

Macroscopic Findings

8 Days After TNBS; 5-Day Treatment

The macroscopic injury observed at 8 days after rats received TNBS was 4.5±0.5. This value indicated that the distal 8 cm of colon contained more than 2 discrete ulcers in an area of inflammation (Fig. 2). In contrast, combination therapy with the NAC plus 5-ASA combination acted synergistically to cause a significant reduction in macroscopic injury as reflected by an injury score of 2.6±0.7; p<0.05. Monotherapy with either NAC or 5-ASA alone caused reduction in macroscopic injury (4.1±0.6 and 3.0±0.7, respectively) but these values did not achieve significance when compared to TNBS alone.

11 Days After TNBS; 8-Day Treatment

Visual evidence of colitis 11 days after TNBS was scored at 4.1±0.2 (Fig. 3). Animal treated with the NAC plus 5-ASA combination for 8 days showed no evidence of mucosal inflammation on ulceration. The colonic mucosa in these animals appeared normal. Monotherapy with either NAC or 5-ASA alone showed mild inflammation and macroscopic injury scores were 1.2±0.2 and 1.8±0.2 respectively.

Microscopic Findings

8 Days After TNBS; 5-Day Treatment

The cumulative or aggregate microscopic colitis injury score for rats examined 8 days after TNBS was 9.4±1.0 which represented moderately severe colitis (Fig. 4). Individual therapy for 5 days with either 5-ASA or NAC alone did not alter the aggregate injury score: 9.0±0.9 and 9.2±1.8, respectively. Therapy with the NAC plus 5-ASA combination, however, caused significant reduction (−44%) in cumulative colitis injury and resulted in a score of 5.0±1.2, P<0.02. Subset analysis of individual histological features, when compared to TNBS alone, indicated that therapy with the NAC plus 5-ASA combination acted synergistically to significantly reduce the degree of epithelial damage, the extent of mucosal ulceration and the amount of mononuclear cell infiltration.

11 Days After TNBS; 8-Day Treatment

Eleven days post TNBS the indices of microscopic colitis were reduced slightly but not significantly from aggregate scores recorded at 8 days after TNBS: 7.0±0.9 vs 9.5±1.0, P>0.1. In contrast to the histological results after 5 days of treatment each of the intraluminal therapies for 8 days caused significant reduction in global measures of colitis (Fig. 5). 5-ASA and NAC each when administered alone caused comparable decrease in aggregate injury of 46 and 53%, respectively. Combination therapy with the NAC plus 5-ASA combination caused a 75% reduction in cumulative colitis injury score: 1.8±0.5 vs 7.0±0.9 TNBS alone; P<0.001. Furthermore, comparisons between monotherapies with either 5-ASA or NAC alone and combination therapy indicated that the NAC plus 5-ASA combination acted synergistically to produce a significantly greater degree of healing than either NAC or 5-ASA alone.

Myeloperoxidase Activity

MPO activity in colonic tissue 11 days after TNBS treatment was elevated greater than 25 fold above values derived from saline treated rats (Fig. 6). In contrast, the NAC plus 5-ASA combination treatment for 8 days reduced MPO activity by 85% to 100.2±12.9 ng/g (P<0.004 vs TNBS 690.2±101.5 ng/g). MPO activity was also reduced by both NAC and 5-ASA alone to levels that were 40% and 32% below TNBS values but these differences did not achieve statistical significance.

Inflammatory Cytokine Expression

Cytokine expression in colonic tissues indicates that elevations in interleukins (IL-1α, IL-1β, IL-4 and IL-6) and TNFα that were induced by TNBS after 11 days were inhibited significantly by 8 day treatment with the NAC plus 5-ASA combination (Table 3). Treatments with either NAC or 5-ASA alone did not significantly suppress levels of cytokine expression induced by TNBS.

### Table 3

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>Treatment Group</th>
<th>Relative O.D. units</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>IL-1A</strong></td>
<td>Saline</td>
<td>0.54 ± 0.07</td>
</tr>
<tr>
<td></td>
<td>TNBS only</td>
<td>1.97 ± 0.3</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA + NAC</td>
<td>0.70 ± 0.08**</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA</td>
<td>1.33 ± 0.18</td>
</tr>
<tr>
<td></td>
<td>TNBS + NAC</td>
<td>1.28 ± 0.16</td>
</tr>
<tr>
<td><strong>IL-1B</strong></td>
<td>Saline</td>
<td>2.25 ± 0.37</td>
</tr>
<tr>
<td></td>
<td>TNBS only</td>
<td>11.77 ± 1.7</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA + NAC</td>
<td>6.03 ± 0.62**</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA</td>
<td>11.49 ± 2.54</td>
</tr>
<tr>
<td></td>
<td>TNBS + NAC</td>
<td>11.88 ± 1.0</td>
</tr>
<tr>
<td><strong>IL-4</strong></td>
<td>Saline</td>
<td>0.27 ± 0.06</td>
</tr>
<tr>
<td></td>
<td>TNBS only</td>
<td>1.76 ± 0.42</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA + NAC</td>
<td>0.82 ± 0.12**</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA</td>
<td>1.40 ± 0.25</td>
</tr>
<tr>
<td></td>
<td>TNBS + NAC</td>
<td>1.20 ± 0.26</td>
</tr>
<tr>
<td><strong>IL-6</strong></td>
<td>Saline</td>
<td>0.18 ± 0.03</td>
</tr>
<tr>
<td></td>
<td>TNBS only</td>
<td>1.21 ± 0.37</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA + NAC</td>
<td>0.46 ± 0.06**</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA</td>
<td>0.79 ± 0.22</td>
</tr>
<tr>
<td></td>
<td>TNBS + NAC</td>
<td>0.76 ± 0.14</td>
</tr>
<tr>
<td><strong>TNFα</strong></td>
<td>Saline</td>
<td>0.41 ± 0.06</td>
</tr>
<tr>
<td></td>
<td>TNBS only</td>
<td>1.51 ± 0.13</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA + NAC</td>
<td>0.7 ± 0.15**</td>
</tr>
<tr>
<td></td>
<td>TNBS + 5ASA</td>
<td>1.25 ± 0.12</td>
</tr>
<tr>
<td></td>
<td>TNBS + NAC</td>
<td>1.04 ± 0.017</td>
</tr>
</tbody>
</table>

**Denotes a P value <0.05 vs TNBS alone.

Data represent the Mean ± SEM of 6-10 observations per condition.

Results of the TNBS experiments of the present study indicate that intraluminal administration of the NAC plus 5-ASA combination to the distal colon of rats with TNBS colitis act synergistically to cause a significant reduction in colonic inflammation and ulceration and acceleration of mucosal healing when compared to either agent used alone. Furthermore, combination therapy with the NAC plus 5-ASA combination caused significantly greater inhibition of myeloperoxidase activity and proinflammatory cytokine...
gene expression in colon of TNBS treated animals than either NAC or 5-ASA alone. Duration of treatment was a
determinant in the effectiveness of antioxidant and anti-in-
flammatory agents on healing of chemically induced colitis.
The NAC plus 5-ASA combination was the only dosing regi-
men that resulted in significant improvement in both macro-
scopic and microscopic measures of colitis after a five day
treatment. In contrast, eight day treatment with NAC alone,
5-ASA alone or the NAC plus 5-ASA combination resulted in
significant improvement in histological features of colitis.
Treatment with the NAC plus 5-ASA combination, however,
causd greater improvement in mucosal injury, inflammation
and epithelial regeneration than NAC or 5-ASA alone. Data
derived from these studies indicate that intraluminal therapy
with the antioxidant NAC plus 5-ASA combination is su-
perior to either agent alone in the treatment of TNBS colitis
and that dual therapy has a synergistic effect in reducing inflam-
mation and promoting mucosal repair.

Colitis induced by intracolic instillation of TNBS mani-
 Maneys many of the histological and clinical features of colonic
inflammatory bowel disease (11). This model of colonic ulceration and transmural inflammation of the mucosa (12)
has been employed to study the pathogenesis of colonic inflammation and to investigate potential treatments of IBD.
In this latter regard previous reports have shown that 5-ASA,
when administered singly by intracolic route to rats, (13)
exerted an anti-inflammatory effect on TNBS colitis. NAC,
administered via drinking water, was observed to increase
colic glutathione stores which were associated with a
reciprocal decrease in the extent of mucosal injury (9).
In support of these observations and the role of ROS generation
in TNBS colitis Luquerio et al reported that glutathione
supplementation improved oxidative damage in TNBS colitis
(14).

Antioxidants, such as NAC, and 5-ASA possess the ability
to scavenge oxygen free radicals, inhibit inducible NO for-
mation and to down regulate nuclear factor kB (NF-kB) activity
(9, 10, 15-17). Furthermore, antioxidants, such as phenyl
N-tetrahydroxytrone, and 5-ASA have been shown to inhibit
cytokine production, including tumor necrosis factor TNFα,
and to retard adhesion molecule expression and B-cell medi-
ated antibody production (10, 18-20) in experimental models
of colitis. Separately and together these agents can, thus, be
evenhanced to moderate immuneocyte (T cell) mediated cyto-
kine elaboration, neutrophil generation of ROS and NO, pro-
staglandin release and to facilitate an environment for unop-
posed cellular and growth factor-mediated tissue repair.
These results show that treatment with the NAC plus 5-ASA
combination caused marked improvement in indices of colitis
and, furthermore, demonstrate prominent features of epitel-
ial repair, and mucosal architectural and glandular restoration.
These data combined with the near normalization of MPO activity and marked reduction in cytokine (ILα, ILb,
II.6) expression indicate that therapy with the NAC plus
5-ASA combination exerts a significantly greater anti-
inflammatory and reparative effect in TNBS colitis than either
5-ASA or NAC when used alone.

Results in the DSS experiments demonstrated that DSS
under these experimental conditions produced mild to mod-
erate colitis. The aggregate microscopic injury score for DSS
treated animals was 5.5±2.0; maximal score 12. Monotherapy
with either 5-ASA or NAC alone caused slight reduction in
aggregate scores to 4.1±0.9 and 4.0±1.1, respectively. These
values were not significantly different from DSS alone. In
contrast, combination therapy with the NAC plus 5-ASA
combination reduced global or aggregate histological injury
score by 67% to 1.8±0.8. Furthermore, the NAC plus 5-ASA
combination caused significant improvement in epithelial
damage when compared to DSS alone. DSS colitis was
associated with elevated CRP levels of 7.0±0.6 mg/ml. CPR
levels were reduced substantially by concurrent therapy with
NAC plus 5-ASA and values were 1.0±0.3 mg/ml. Similar
reductions in CRP levels were observed with either 5-ASA or
NAC alone. DSS treatment caused marked elevation in cytokine
gene expression for IL 1a and II 1b and these values represented a
6.8 and 12.1 fold increase, respectively, in gene
expression of these cytokines when measured above control
values.

NAC and 5-ASA, alone or in combination, substantially
reduced DSS-induced IL1a and II1b gene expression by
55-90% to levels that approximated control values. Conclu-
sions from these data indicate that intraluminal therapy with
the NAC plus 5-ASA combination caused significant amelio-
ration of mucosal injury induced by DSS. Combination treat-
ment with the NAC plus 5-ASA combination results were
associated with substantial reduction in serum CRP levels and
proinflammatory cytokine gene expression.

Current considerations of the pathogenesis of mucosal
inflammation in IBD involve a number of steps from antigen
presentation and processing by macrophages to amplification
of T cell activation and differentiation and cytokine produc-
tion (1). In addition, inflammatory cells, including granulo-
cyes and mononuclear cells, are recruited to the mucosa in a
highly coordinated fashion. Once present in the inflamed
mucosa, tissue injury is enhanced by neutrophil production of
reactive oxygen species such as superoxide and an increase in
the expression of the inducible isoform of NO synthase
(iNOS) (5). Mucosal healing is thought to occur, in part, by
reduction in injurious and proinflammatory substances and,
also, by local liberation of growth factors which facilitate
cellular restitution and repair (2). Although several experi-
mental approaches have been employed that suggest the
importance of enhanced production of superoxide and nitric
oxide in the pathogenesis of IBD, inconsistent results have
this issue unresolved (5). The beneficial effect superoxide
dismutase (SOD) treatment in experimental models of colitis
(21-23) has shown limited effect in humans with IBD (24).
In addition, iNOS inhibition has shown variable results in
experimental models of IBD (25-27). However, recent studies
using gene-targeted mice suggest a dominant role of iNOS-
derived NO in a murine model of dextran sulfate sodium
(DSS) colitis (5). The antioxidants NAC and phenyl N-tet-
butylintrone have been shown to be effective in reducing the
injurious consequences of TNBS colitis in rats (9) and DSS
colitis in mice (10), respectively. Antioxidant therapy has also
been demonstrated to suppress colonic iNOS activity and to
decrease NF-kB DNA-binding activity in experimental coli-
tis (10) and man (16).

In conclusion, our results showed that treatment of TNBS-
induced colitis with the NAC plus 5-ASA combination was
superior to either 5-ASA or NAC when used alone in reducing
colonic inflammation and in promoting mucosal repair. In
addition, combination therapy with the NAC plus 5-ASA
combination acted synergistically to result in a significant
reduction in MPO activity and proinflammatory cytokine
gene expression.

**UTILITY**

The present invention provides a method for the treatment of a mammal subject (including humans) afflicted with
inflammatory diseases, and in particular, inflammatory bowel
diseases. The present invention has several advantages over
current therapies. As demonstrated herein, combination
therapy with 5-ASA plus an antioxidant is synergistically superior to either agent alone in controlling mucosal inflammation. It is envisioned that such combination therapy would be capable of being delivered by oral route, employing existing coating technologies, to sites of inflammation. In addition, such combination therapy would be amenable to local therapy in the distal colon and rectum by enema or suppository. Further description regarding deliver methods and dosing systems and protocols is discussed below.

The combination of at least two active anti-inflammatory agents into a single delivery system as described herein provides greater clinical efficacy, development of a new pharmacological approach for treating mucosal inflammatory conditions, and a reduction in the need for other potentially toxic and expensive anti-inflammatory agents. The chemical coupling of 5-ASA to an antioxidant substance can provide a further pharmacological approach to the treatment of mucosal inflammatory conditions such as IBD.

The 5-ASA used in the composition of the present invention would be provided as the free acid, or as a pharmaceutically acceptable salt or ester, for example as described in U.S. Pat. No. 5,013,727, the entirety of which is hereby expressly incorporated by reference herein.

While it is contemplated that applications of the invention would be principally of treating mucosal inflammatory conditions associated with inflammatory bowel diseases, such as ulcerative colitis, Crohn's disease and Behcet's disease, the invention would also have application to other disorders of the gastrointestinal tract such as radiation and infective enteritis, ischemic injury to the gastrointestinal tract, infections, caustic agent-induced gastrointestinal injury, hemorrhagic rectal ulcers, ileum pouchitis, ischemic enteritis and drug-induced colitis, mucous colitis, pseudomembranous enterocolitis, non-specific colonic ulcers, colangitis colitis, cathartic colon, ulcerative proctitis, idiopathic diffuse ulcerative non-granulomatous enteritis, non-steroidal anti-inflammatory drug-induced inflammations, celiac sprue and the like. Furthermore, advances in our understanding of the pathogenesis of gastrointestinal malignancies suggest a role for prostaglandins generated by the cyclooxygenase enzymes to have a role in neoplasia. It has been well established that nonspecific and specific cyclooxygenase inhibitors can reduce the propensity to neoplasia and malignancy in experimental models and in human subjects. Full appreciation of the role of prostaglandins in the cyclooxygenase system in the pathogenesis of gastrointestinal malignancy is not complete. However, it is conceivable that therapies which combine inhibition prostaglandin synthesis through the cyclooxygenase enzymes and antioxidant therapies may have a beneficial role in preventing gastrointestinal malignancies such as colorectal cancer.

The term "inflammation" as used herein is meant to include reactions of both the specific and non-specific defense systems. A specific defense system reaction is a specific immune system reaction to an antigen. Examples of a specific defense system reaction include the antibody response to antigens such as rubella virus, and delayed-type hypersensitivity response mediated by T-cells (as seen, for example, in individuals who test "positive" in the Mantoux test).

A non-specific defense system reaction is an inflammatory response mediated by leukocytes incapable of immunological memory. Such cells include granulocytes, macrophages, neutrophils, for example. Examples of a non-specific defense system reaction include the immediate swelling at the site of a bee sting, the reddening and cellular infiltrate induced at the site of a burn and the collection of PMN leukocytes at sites of bacterial infection (e.g., pulmonary infiltrates in bacterial pneumonia, pus formation in abscesses).

Although the invention is particularly suitable for cases of acute inflammation, it also has utility for chronic inflammation. Types of inflammation that can be treated with the present invention include diffuse inflammation, traumatic inflammation, immunosuppression, toxic inflammation, specific inflammation, reactive inflammation, parenchymatous inflammation, obliterative inflammation, interstitial inflammation, croupous inflammation, and focal inflammation.

A therapeutically effective amount of a composition of the present invention refers to an amount which is effective in controlling, treating or moderating the inflammatory response. The terms "controlling," "treating" or "moderating" are intended to refer to all processes wherein there may be a slowing, interrupting, arresting, or stopping of the progression of the disease and does not necessarily indicate a total elimination of all disease symptoms.

The term "therapeutically effective amount" is further meant to define an amount resulting in the improvement of any parameters or clinical symptoms characteristic of the inflammatory response. The actual dose will be different for the various specific molecules, and will vary with the patient's overall condition, the seriousness of the symptoms, and counter indications.

As used herein, the term "subject" or "patient" refers to a warm blooded animal such as a mammal which is afflicted with a particular inflammatory disease state. It is understood that guinea pigs, dogs, cats, rats, mice, horses, cattle, sheep, goats, pigs, llamas, and humans are among the examples of animals within the scope of the meaning of the term.

A therapeutically effective amount of the compound used in the treatment described herein can be readily determined by the attending diagnostician, as one skilled in the art, by the use of conventional techniques and by observing results obtained under analogous circumstances. In determining the therapeutically effective dose, a number of factors are considered by the attending diagnostician, including, but not limited to: the species of mammal; its size, age, and general health; the specific disease or condition involved; the degree of or involvement or the severity of the disease or condition; the response of the individual subject; the particular compound administered; the mode of administration; the bioavailability characteristic of the preparation administered; the dose regimen selected; the use of concomitant medication; and other relevant circumstances.

A therapeutically effective amount of the compositions of the present invention will generally contain sufficient active ingredient (i.e., the antioxidant and 5-ASA) to deliver from about 0.1 μg/kg to about 6000 μg/kg (weight of active ingredient/body weight of patient). Preferably, the composition will deliver at least 1.0 μg/kg to 1000 μg/kg, and more preferably at least 1 mg/kg to 100 mg/kg, although each dose of the composition may be more or less than these amounts. For example, the daily dose for an adult may be in the range of about 10 μg to 1000 μg/kg, preferably in the range of about 20 μg to 300 μg/kg, especially in the range of 50 μg/kg to 200 μg/kg. Also see U.S. Pat. No. 5,013,727 which is incorporated by reference herein.

Practice of the method of the present invention comprises administering to a subject a therapeutically effective amount of the composition described herein, in any suitable systemic or local formulation, in an amount effective to deliver the dosages listed above. The dosage can be administered on a one-time basis, or (for example) from one to five times per day or once or twice per week, or continuously via a venous drip, depending on the desired therapeutic effect.
As noted, preferred amounts and modes of administration are able to be determined by one skilled in the art. One skilled in the art of preparing formulations can readily select the proper form and mode of administration depending upon the particular characteristics of the compound selected, the disease state to be treated, the stage of the disease, and other relevant circumstances using formulation knowledge known in the art, for example, in Remington’s Pharmaceutical Sciences, latest edition, Mack Publishing Co.

Pharmaceutical compositions can be manufactured utilizing techniques known in the art. Typically the therapeutically effective amount of the compound will be admixed with a pharmaceutically acceptable carrier.

The compounds or compositions of the present invention may be administered by a variety of routes, for example, orally, intrarectally or parenterally (i.e., subcutaneously, intravenously, intramuscularly, intraperitoneally, or intratra
cinally).

For oral administration, the compounds can be formulated into solid or liquid preparations such as capsules, pills, tablets, lozenges, melts, powders, suspensions, or emulsions. Solid unit dosage forms can be capsules of the ordinary gelatin type containing, for example, surfactants, lubricants and inert fillers such as lactose, sucrose, and cornstarch or they can be sustained release preparations.

In another embodiment, the compounds of this invention can be tabletted with conventional tablet bases such as lactose, sucrose, and cornstarch in combination with binders, such as avicel, cornstarch or gelatin, disintegrating agents such as potato starch or alginic acid, and a lubricant such as stearic acid or magnesium stearate. Liquid preparations are prepared by dissolving the active ingredient in an aqueous or non-aqueous pharmaceutically acceptable solvent which may also contain suspending agents, sweetening agents, flavoring agents, and preservative agents as are known in the art.

For parenteral administration, the compounds may be dissolved in a physiologically acceptable pharmaceutical carrier and administered as either a solution or a suspension. Illustrative of suitable pharmaceutical carriers are water, saline, dextrose solutions, fructose solutions, ethanol, or oils of animal, vegetative, or synthetic origin. The pharmaceutical carrier may also contain preservatives, and buffers as are known in the art.

The compounds of this invention can also be administered topically. This can be accomplished by simply preparing a solution of the compound to be administered, preferably using a solvent known to promote transdermal absorption such as ethanol or dimethyl sulfoxide (DMSO) with or without other excipients. Preferably topical administration will be accomplished using a patch either of the reservoir and porous membrane type or of a solid matrix variety.

As noted above, the compositions can also include an appropriate carrier. For topical use, any of the conventional excipients may be added to formulate the active ingredients into a lotion, ointment, powder, cream, spray, or aerosol. For surgical implantation, the active ingredients may be combined with any of the well-known biodegradable and bioerodible carriers, such as polylactic acid and collagen formulations. Such materials may be in the form of solid implants, sutures, sponges, wound dressings, and the like. In any event, for local use of the materials, the active ingredients usually be present in the carrier or excipient in a weight ratio of from about 1:1,000 to 1:20,000, but are not limited to ratios within this range. Preparation of compositions for local use are detailed in Remington’s Pharmaceutical Sciences, latest edition, (Mack Publishing).

Additional pharmaceutical methods may be employed to control the duration of action. Increased half-life and controlled release preparations may be achieved through the use of polymers to conjugate, complex with, or absorb the composition described herein. The controlled delivery and/or increased half-life may be achieved by selecting appropriate macromolecules (for example, polysaccharides, polymers, polyamino acids, homopolymers polystyrylpyrrolidone, ethylenevinylacetate, methylecellulose, or carbomethylecellulose, and acrylamides such as N-(2-hydroxypropyl) methacrylamide, and the appropriate concentration of macromolecules as well as the methods of incorporation, in order to control release.

Another possible method useful in controlling the duration of action by controlled release preparations and half-life is incorporation of the glycospaltenepptide molecule or its functional derivatives into particles of a polymeric material such as polyesters, polyamides, polyamino acids, hydrogels, poly(lactic acid), ethylene vinylacetate copolymers, copolymer micelles of, for example, PEG (polyethylene glycol) and poly(l-ascaptamid).

Alternatively, it is possible to entrain the compositions in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization (for example, hydroxyethylcellulose or gelatin-microcapsules and polyvinyl (methylmethacrylate) microcapsules, respectively), in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles, and microcapsules), or in macroemulsions. Such techniques are disclosed in the latest edition of Remington’s Pharmaceutical Sciences.

U.S. Pat. No. 4,789,734 describe methods for encapsulating compositions in liposomes and is hereby expressly incorporated by reference herein. Essentially, the material is dissolved in an aqueous solution, the appropriate phospholipids and lipids added, along with surfactants if required, and the material dialyzed or sonicated, as necessary. A review of known methods is by G. Gregoriadis, Chapter 14, “Liposomes”, Drug Carriers in Biology and Medicine, pp. 287-341 (Academic Press, 1979). Microspheres formed of polymers or proteins are well known to those skilled in the art, and can be tailored for passage through the gastrointestinal tract directly into the blood stream. Alternatively, the agents can be incorporated and the microspheres, or composite of micro
spheres, implanted for slow release over a period of time, ranging from days to months. See, for example, U.S. Pat. Nos. 4,906,474; 4,925,673; and 3,625,214 which are expressly incorporated by reference herein.

When the composition is to be used as an injectable material, it can be formulated into a conventional injectable carrier. Suitable carriers include biocompatible and pharmaceutically acceptable phosphate buffered saline solutions, which are preferably isotonic.

For reconstitution of a lyophilized product in accordance with this invention, one may employ a sterile diluent, which may contain materials generally recognized for approximating physiological conditions and/or as required by governmental regulation. In this respect, the sterile diluent may contain a buffering agent to obtain a physiologically acceptable pH, such as sodium chloride, saline, phosphate-buffered saline, and/or other substances which are physiologically acceptable and/or safe for use. In general, the material for intravenous injection in humans should conform to regulations established by the Food and Drug Administration, which are available to those in the field.

The pharmaceutical composition may also be in the form of an aqueous solution containing many of the same substances as described above for the reconstitution of a lyophilized product.
The compounds can also be administered as a pharmaceutically acceptable acid- or base-addition salt, formed by reaction with inorganic acids such as hydrochloric acid, hydrobromic acid, perchloric acid, nitric acid, thiocyanic acid, sulfuric acid, and phosphoric acid, and organic acids such as formic acid, acetic acid, propionic acid, glycolic acid, lactic acid, pyruvic acid, oxalic acid, malonic acid, succinic acid, maleic acid, and fumaric acid, or by reaction with an inorganic base such as sodium hydroxide, ammonium hydroxide, potassium hydroxide, and organic bases such as mono-, di-, trialkyl and aryl amines and substituted ethanolamines.

As mentioned above, the compounds of the invention may be incorporated into pharmaceutical preparations which may be used for therapeutic purposes. However, the term “pharmaceutical preparation” is intended in a broader sense herein to include preparations containing a 5-ASA/antioxidant composition in accordance with this invention, used not only for therapeutic purposes but also for reagent or diagnostic purposes as known in the art, or for tissue culture. The pharmaceutical preparation intended for therapeutic use should contain a “pharmacologically acceptable” or “therapeutically effective amount” of the composition, i.e., that amount necessary for preventative or curative health measures. If the pharmaceutical preparation is to be employed as a reagent or diagnostic, then it should contain reagent or diagnostic amounts of a 5-ASA/antioxidant combination.

All references, patents and patent applications cited herein are hereby incorporated herein in their entirety by reference.

The present invention is not to be limited in scope by the specific embodiments described herein, since such embodiments are intended as but single illustrations of one aspect of the invention and any functionally equivalent embodiments are within the scope of this invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and accompanying drawings.

CITED REFERENCES


What is claimed is:

1. A method of reducing macroscopic injury in a colonic mucosa of a mammal, comprising: identifying a mammal in need thereof, and administering to the mammal a therapeutically effective amount of 5-aminosalicylic acid, or a pharmaceutically acceptable salt thereof, and a therapeutically effective amount of N-acetylcysteine, or a pharmaceutically acceptable salt thereof, wherein the macroscopic injury to the colonic mucosa of the mammal is reduced.

2. The method of claim 1, wherein the mammal is a human.

3. The method of claim 1, wherein the 5-aminosalicylic acid or a pharmaceutically-acceptable salt thereof, and the N-acetylcysteine or a pharmaceutically-acceptable salt thereof, are together disposed in a dosage form further comprising a pharmaceutically acceptable carrier or excipient.

4. The method of claim 3, wherein the dosage form is administered to the mammal at a rate which comprises providing the dosage form daily for 8 days.

5. The method of claim 1, wherein the 5-aminosalicylic acid or a pharmaceutically-acceptable salt thereof is administered at a dose of between 1 mg/kg to 100 mg/kg, and wherein the N-acetylcysteine or a pharmaceutically-acceptable salt thereof is administered at a dose of between 1 mg/kg to 100 mg/kg.

6. A method of reducing macroscopic injury in a colonic mucosa of a mammal, comprising: identifying a mammal in need thereof, and administering to the mammal 5-aminosalicylic acid, or a pharmaceutically-acceptable salt thereof, at a dose of between 1 mg/kg to 100 mg/kg, and N-acetylcysteine, or a pharmaceutically-acceptable salt thereof, at a dose of between 1 mg/kg to 100 mg/kg, wherein the macroscopic injury to the colonic mucosa of the mammal is reduced.

7. The method of claim 6, wherein the 5-aminosalicylic acid or a pharmaceutically-acceptable salt thereof, and the N-acetylcysteine or a pharmaceutically-acceptable salt thereof, are together disposed in a dosage form further comprising a pharmaceutically acceptable carrier or excipient.

8. The method of claim 6, wherein the mammal is a human.

9. The method of claim 7, wherein the dosage form is administered to the mammal at a rate which comprises providing the dosage form daily for 8 days.

10. A method of reducing macroscopic injury in a colonic mucosa of a mammal, comprising: identifying a mammal in need thereof, and administering to the mammal a dosage form comprising a therapeutically effective amount of 5-aminosalicylic acid, or a pharmaceutically-acceptable salt thereof, and a therapeutically effective amount of N-acetylcysteine, or a pharmaceutically-acceptable salt thereof, wherein the macroscopic injury to the colonic mucosa of the mammal is reduced, and wherein the dosage form is administered to the mammal at a rate which comprises providing the dosage form daily for 8 days.

11. The method of claim 10, wherein the dosage form comprises 5-aminosalicylic acid or a pharmaceutically-acceptable salt thereof at a dose of between 1 mg/kg to 100 mg/kg, and N-acetylcysteine or a pharmaceutically-acceptable salt thereof at a dose of between 1 mg/kg to 100 mg/kg.

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